

## **Identification and Biological Characterization of Angiogenic and Tumor Growth Inhibitors derived from *Sinica cetorhinus maximum* Cartilage**

**Binghua Jiao\***, Jianhe Chen, Weimin Miao, Lianghua Wang, Yuping Zhu and Huinan Miao

Department of Biochemistry and Molecular Biology, Faculty of Basic Medical Sciences, Second Military Medical University, Shanghai 200433, P.R. China. Tel. (021)-65493936, Fax. (021)-65334344

\* Author to whom correspondence should be addressed; E-mail: jiaobh@uninet.com.cn

Received: 25 March 2003 / Accepted: 31 January 2004 / Published: 25 February 2004

---

**Abstract:** Shark (*Sinica cetorhinus maximum*) cartilage was extracted in 1 mol/L Gu-HCl guanidine. Two purified active proteins with apparent molecular weights of  $15.2 \times 10^3$  Da and  $8.0 \times 10^3$  Da (designated as Sp15 and Sp8, respectively) were obtained through ultrafiltration and Superdex 75 chromatography. The activities of the samples were studied in terms of their potential inhibition of vascular endothelial cell growth *in vitro*, of angiogenesis both in rabbit cornea and chick embryo chorioallantoic membrane (CAM) assay models *in vivo*, and of growth of transplanted S180 sarcoma in mice *in vivo*. The results showed that Sp15 expressed a typical lysozymatic activity up to 223,000 U/mg and its N-terminus was highly homologous to lysozymes of various mammalian origins. Sp15 exhibited a strong anti-angiogenic activity only *in vitro*, whereas Sp8 shared this effect both *in vitro* and *in vivo*. Both Sp15 and Sp8 provided an effective anti-tumor activity in mice bearing transplanted S180 sarcoma. These results suggest that Sp15 is a shark cartilage-derived lysozyme that participates in the defense to bacterial invasion to the body, while Sp8 is an angiogenic inhibitor that mediates at least part of the anti-tumor activity associated with shark cartilage probably through the inhibition of tumor-induced angiogenesis.

**Keywords:** Shark cartilage, lysozyme, angiogenesis, anti-tumor activity.

---

## Introduction

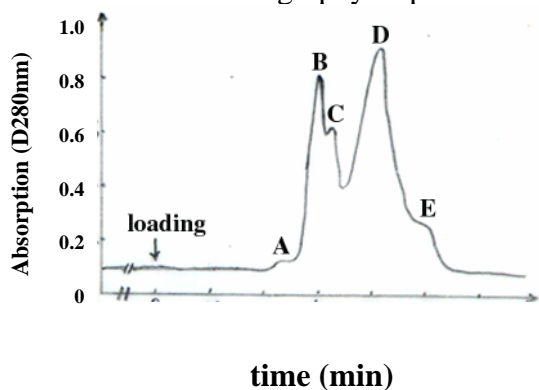
Neovascularization is highly correlated with the rapid growth and metastasis of solid tumors. Various angiogenic inhibitors including angiostatin, endostatin, THP-470, metalloproteinase inhibitors and Bufotanine as the therapeutic intervention for angiogenesis are being tested for the treatment of cancers [1]. Recently, Genentech developed a humanized monoclonal antibody to VEGF (Bevacizumab, Avastin); it is now under Phase III clinical trials for the treatment of colorectal, breast, and non-small cell of the lung cancer [2]. Several studies have indicated that shark cartilage is endowed with effective angiogenic inhibition and anti-tumor activities [3-5], the molecular mechanisms for which remain unclear however. This investigation describes the separation and purification of shark cartilage-derived proteins, and the screening for active molecules *via* tests for *in vitro* vascular endothelial cell proliferation-inhibition, *in vivo* angiogenic inhibition and *in vivo* anti-tumor activity on mice-bearing transplanted sarcoma 180 (S180).

## Results and Discussion

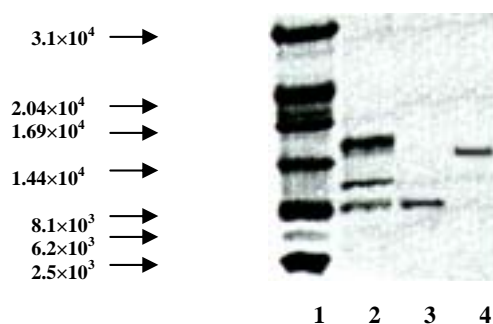
### *Purification and identification of Sp15 and Sp8*

Previous studies on shark cartilage-derived active protein(s) [3-7] indicated that the molecular weights of the main active components seemed to be less than  $3.0 \times 10^4$  Da, which prompted us to investigate the composition and activities of lower molecular mass species. Several extraction protocols including ultrafiltration and column chromatography have been attempted to isolate and identify the active fractions from shark cartilage (see the part of Experimental), and our data showed that Sp15 was the predominant component in shark cartilage, accounting for approximately more than 50% w/w in extracts, whereas Sp8 accounted for ~20%. Superdex 75 chromatography yielded the highly purified Sp15 and Sp8 in the fractions designated B and D, respectively (Figure 1). A Tricine-SDS-PAGE was used to determine their apparent molecular masses as  $1.52 \times 10^4$  Da for Sp15 and  $8.3 \times 10^3$  Da for Sp8, respectively (Figure 2).

**Figure 1** Superdex 75 column chromatography of proteins derived from shark cartilage



Fraction B: lysozyme-like molecule (Sp15); D: angiogenic inhibitor (Sp8).

**Figure 2.** Tricine-SDS-PAGE of proteins derived from shark cartilage

1: protein marker; 2: shark cartilage proteins extracted by Gu-HCl and after ultrafiltration; 3: purified Sp8 from Superdex 75 chromatography; 4: purified Sp15 from Superdex 75

#### *Determination of the N-terminus and lysozymatic activity of Sp15*

The N-terminus of Sp15, as determined by an Edman degradation method, was YTYQKHELARVLQSKGLD(X)QYG (X: unclear). The result showed that Sp15 is a newly identified protein with a high degree of homology to lysozymes derived from human, rat, mouse, dog, cat and rabbit, respectively. The specific lysozymatic activity of Sp15 was determined in a *Micrococcus lysodeikticus* lytic assay with egg white lysozyme as the control and it was as high as 223,000 U/mg, suggesting that it was a typical lysozyme derived from shark cartilage.

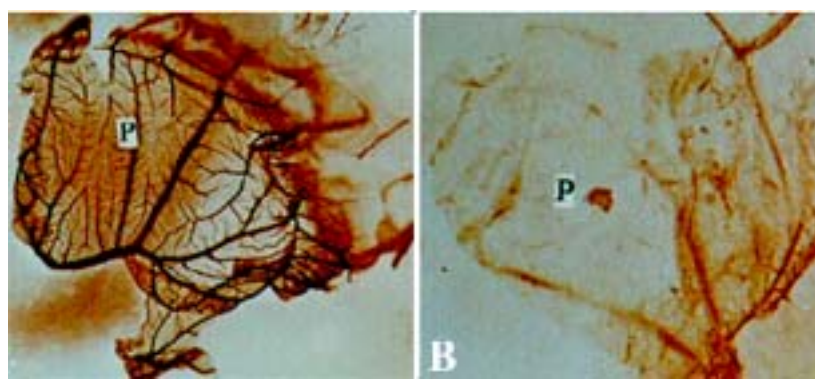
#### *Inhibition of endothelial cell proliferation*

The inhibition of proliferation by Sp15 and Sp8 on cultivated rat pulmonary capillary and bovine aortic endothelial cells was investigated (data not shown). Results showed that the inhibitory effect of both Sp15 and Sp8 was dose-dependent within the particular range of the concentrations applied. The highest inhibitory rate at 75.0% was reached for Sp15 at 10.0 µg/mL, while it was 20.0% for Sp8 at 40.0 µg/mL (data not shown). Although the inhibition by Sp8 seems to be weaker than that by Sp15, the inhibition by Sp8 was prolonged: it usually took 2-3 d for *in vitro* cultivated endothelial cells to form a monolayer, but when Sp8 is added to the medium even in a lower concentration (5.0 µg/mL), such a feature could not be seen even after 1 week's cultivation. In contrast, endothelial cells would grow normally and form monolayers as before when Sp8 was removed from the medium. The results also indicated that both Sp15 and Sp8 display no inhibition of proliferation to cultivated tumor and transformed cell lines, including B16 melanoma and L929 fibroblast.

*Inhibition of angiogenesis of Sp8 in vivo*

We observed Sp8-mediated angiogenic inhibition on two models, rabbit cornea and chick embryo CAM models, respectively. Both have been used extensively in the angiogenic inhibitor research. The results showed that Sp8 exhibited a strong inhibitory effect up to a rate of 79.0% on the CAM model (Figure 3) and 72.7% on rabbit cornea model (Table 1, Figure 4) at the dose of 40.0  $\mu\text{g}$ , respectively. No obvious dose response relationship has been observed at the range of 20.0~160.0  $\mu\text{g}$ . The results indicate that Sp8 is a potent angiogenic inhibitor present in shark cartilage. In the same test model, Sp15 showed no angiogenic inhibition *in vivo*.

**Figure 3.** Sp8 inhibits angiogenesis in CAM



Sp8 (left); B: control (right); P: EVA.

**Table 1** Sp8 inhibits LPS-induced angiogenesis in rabbit cornea model (n=4)

Sample <sup>a</sup>	Angiogenesis			Inhibition rate (%) <sup>c</sup>
	Length (mm)	Clock No.	Area (mm <sup>2</sup> ) <sup>b</sup>	
LPS	5.0±0.3	5.6	46.90±4.32	0
Sp8+LPS	1.2±0.2	4.0	12.82±3.13	72.7**

a: LPS 140  $\mu\text{g}$ , Sp8 40  $\mu\text{g}$ ;

b: area (S) =  $C/12 \times 3.1416 \times [r^2 - (r-L)^2]$  (C: Clock number; r: Radius of cornea; L: Length);

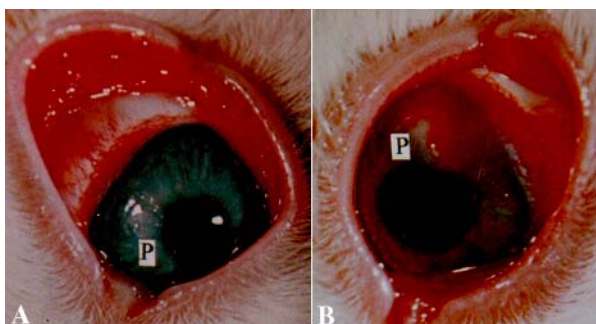
c: Inhibition rate =  $[(S_{\text{control}} - S_{\text{sample}}) / S_{\text{control}}] \times 100\%$ ;

\*\* $p < 0.01$

*Anti-tumor activity of Sp15 and Sp8 in vivo*

The anti-tumor activities of Sp15 and Sp8 were observed on mice bearing transplanted S180 sarcoma. C57/BL6 mice were transplanted  $2 \times 10^7$  S180 cells subcutaneously and different doses of Sp15 and Sp8 in three groups each were given intraperitoneally at day 2, with PBS and cyclophosphamide (CTX) as controls. The result showed that 2.0 mg/kg of Sp15 exhibited no inhibition to tumor growth, while doses of 4.0 and 8.0 mg/kg yielded inhibitory rates of 23.3% ( $p < 0.05$ ) and 33.3% ( $p < 0.01$ ), respectively. Only 10% inhibition was seen in the control group of egg white lysozyme at 20mg/kg (Table 2). Furthermore, mice generally died at day 11 after they were inoculated  $5 \times 10^7$  S180 sarcoma cells by peritoneal injection. When these mice bearing S180 ascites were treated with 8 mg/kg of Sp15, their mean life-span was prolonged up to day 17.2 ( $p < 0.01$ ), while those treated with 20 mg/kg of egg white lysozyme died at day 15.2 (data not shown). These data demonstrate that Sp15 has more effective anti-tumor activity in this model than that of egg white one.

**Figure 4.** Sp8 inhibits LPS-induced angiogenesis on rabbit cornea



A: Sp8+LPS; B: LPS control; P: EVA.

Sp8 also inhibited significantly the rapid growth of the tumor mass at the rate of 21.1, 54.4 and 56.7%, respectively, corresponding to the doses of 5.0, 10.0 and 20.0  $\mu\text{g}/\text{kg}$ , respectively (Table 2). Furthermore, the application of Sp8 to this model inhibited nearly 100% of tumor-related skin ulcer formation (data not shown).

No obvious systemic toxicities such as weight loss were observed upon the treatment of Sp15 and Sp8.

## Conclusions

Cartilage is one the few tissues containing biological inhibitors of angiogenesis [7]. Various researchers have isolated and characterized several active fractions from shark cartilage while no detailed physico-chemical properties were provided [4-5, 8]. Cartilage-derived angiogenic

inhibitors exhibited a significant anti-tumor activity *in vivo*. Recently, Neovastat, a naturally occurring inhibitor of angiogenesis derived from marine cartilage (dogfish), has been tested in Phase II clinical trials in non-small cell lung cancer and in renal cell carcinoma. It is currently undergoing Phase III clinical trials for the treatment of refractory renal cell carcinoma and nonresectable small cell lung cancer in addition to a Phase II pivotal clinical trials for the treatment of recurrent multiple myeloma. Tumor regressions were seen in some patients and it has been shown highly safe by repeated administration [8-10].

Table 2. Sp15 and Sp8 inhibit the growth of S180 solid sarcoma transplanted in mice<sup>a</sup>

Sample	Dosage (mg/kg)	Weight of tumor (g±x)	Inhibitory rate <sup>b</sup> (%)
Sp15	2.0	0.90±0.09	0
	4.0	0.69±0.15	23.3*
	8.0	0.60±0.11	33.3**
Ely <sup>c</sup>	20.0	0.81±0.18	10.0*
Sp8	0.050	0.71±0.12	21.1*
	0.100	0.41±0.11	54.4**
	0.200	0.39±0.05	56.7**
CTX <sup>d</sup>	3.0	0.62±0.13	31.0**
PBS	-	0.90±0.15	-

a: each dose was tested in 20 mice;

b: inhibitory rate = [(tumor weight of control - tumor weight of experiment) / tumor weight of control] × 100%;

c: Ely: egg white lysozyme;

d: CTX (cyclophosphamide).

\*  $p < 0.05$ ; \*\*  $p < 0.01$ .

This investigation indicates that shark cartilage contains an active molecule with a weight of  $8.3 \times 10^3$  Da (Sp8) which exhibits a stronger inhibition on the proliferation of *in vitro* cultivated vascular endothelial cells as well as a predominant inhibition of angiogenesis in the rabbit cornea and chick embryo CAM models *in vivo*. Based on the mechanisms of new capillary formation, the proliferation of the endothelial cells is the most important prerequisite. We are now investigating if Sp8 is involved in the inhibition of other processes of neovascularization including the inhibition of matrix metalloproteinase activity and the migration of the endothelial cells. This paper shows also that Sp8 significantly inhibits the growth of S180 tumor mass in mice model and of tumor-related

ulcer formation. The anti-tumor activity of Sp8 to other tumor cell lines and its pharmacodynamics and long-term toxicities by repeated injections are also being investigated in our laboratory.

Besides the angiogenic inhibitors, shark cartilage contains also an abundant amount of lysozyme-like substance (Sp15). Lysozyme has been studied for its anti-tumor activity for more than forty years (see the review of Sava G, et al [11]). The present study showed that Sp15 expressed a higher inhibitory activity against the growth of S180 sarcoma on tumor-bearing mice, and could also prolong the life-span of the mice. The results indicated that Sp15, a shark cartilage derived lysozyme, may contribute, at least partly, to the anti-tumor activity displayed by shark cartilage.

## Experimental

### General

Fresh mandibular cartilage was obtained from shark (*sinica cetorhinus maximum*) captured from the deep sea in Bohai, China. Experimental animals including SD rats, C57/BL6 mice and New Zealand rabbits were provided by Shanghai Laboratory Animals Center. Rat pulmonary capillary endothelial (RCE) and new-born bovine aortic endothelial (BAE) cells were prepared and cultivated according to the methods described by Chen, et al [12] and DeClerck and Lung [13], respectively. Superdex G-75 was purchased from Pharmacia; Gu-HCl (A.R.) from Jassen; MES and Tricine from Amersco; Lower molecular weight marker and non-radioactive cell-proliferation detecting kit from Promega; Ultrafilter and its membrane and PVDF membrane from Millipore.

### Extraction and Purification

Sp15 and Sp8 were extracted and purified as follows: 2,000g of cartilage was chopped into pieces before homogenization, and extracted in 10 L of 0.02 mol/L MES, pH 6.0, containing 1 mol/L Gu-HCl for 120 h at room temperature. The extracts were centrifuged at  $1.1 \times 10^4$  xg for 1 h at 4°C and filtered through multiple layers of cheesecloth. The filtered extracts, approximately 8 L in volume, were firstly filtered on Millipore membrane PTTK  $3.0 \times 10^4$  and then through membrane PLAC  $1.0 \times 10^3$ . The extracts were subjected to Superdex 75 (1.0x 60cm) to obtain the highly purified Sp15 and Sp8.

### Analytical

A Tricine-SDS-PAGE technique was applied for the determination of the apparent molecular masses of the active proteins using the procedure of Schagger, et al [14]. Sp15 was run on Tricine-SDS-PAGE and electrically transferred to PVDF membrane at 150mA in CAPS buffer containing 10% methanol at pH 11.0. PVDF membrane was stained with Commassie blue bright R-250. The band corresponding to Sp15 was cut off with a clean blade and rinsed with distilled water. The N-

terminal sequence of Sp15 was determined by an automated Edman degradation on Applied Biosystem Model PROCICE protein sequencer.

The lysozymatic activity of Sp15 was determined by a standard *Micrococcus lysodeikticus* lytic assay with egg white lysozyme as the control.

A MTT test was used for determining the inhibition of endothelial cell proliferation. Inhibition of angiogenesis on rabbit cornea model was performed according to Langer, et al [15]: lipopolysaccharide (LPS, endotoxin, 140 µg) isolated from *Salmonella abortus equi* was mixed with Sp8 (40 µg) into ethylene-vinyl acetate (EVA) to prepare the sustained releasing compound and then transplanted into rabbit cornea; 10 d later, the result of angiogenic inhibition was recorded. Chick embryo chorioallantoic membrane (CAM) model was also used to assess the inhibition of angiogenic effect of Sp8 according to Taylor, et al [16]: Sp8 was mixed into EVA to prepare the sustained releasing compound and transplanted into the capillary-free area of CAM of chick embryo at 9 d old of age; 3 d later, it was taken out and the result of the inhibition were recorded. *In vivo* tumor growth inhibition test was performed as the routine procedure.

## References and Notes

1. Hahnfeldt, P., Panigrahy, D., Folkman, J., Hlatky, L. Tumor development under angiogenic signaling: dynamical theory of tumor growth, treatment response, and postvascular dormancy. *Cancer Res.* **1999**, *59*, 4770-4775.
2. Ferrara, N. Role of vascular endothelial growth factor in physiologic and pathologic angiogenesis: therapeutic implications. *Semin Oncol.* **2002**, *29* (6 Suppl 16), 10-4.
3. Wang, L., Jiao, B., Miao, H. Isolation, purification and physico-chemical characterization of an angiogenetic inhibitor derived from *Sinica cetorhinus maximum* cartilage. *Pharm Biotech.* **1997**, *4*, 17-21.
4. Langer, R., Brem, H., Falterman, K., Klein, M., Folkman, J. Isolation of a cartilage factor inhibits tumor neovascularization. *Science* **1976**, *193*, 70-72.
5. Oikawa, T., Ashino-Fuse, H., Shimamura, M., Koide U, Lwaguchi T. A novel angiogenic inhibitor derived from Japanese shark cartilage (I): Extraction and estimation of inhibitory activities toward tumor and embryonic angiogenesis. *Cancer Lett.* **1990**, *51*, 181-186.
6. Liang, J.H, Wong. K.P. The characterization of angiogenesis inhibitor from shark cartilage. *Adv Exp Med Biol.* **2000**, *476*, 209-223.
7. Brem, H., Folkman, J. Inhibition of tumor angiogenesis mediated by cartilage. *J Exp Med.* **1975**, *141*, 427-439.
8. Dupont, E., Brazeau, P., Juneau C. Extracts of shark cartilage having an antiangiogenic activity and an effect on tumor regression; process of making thereof. United States Patent 5618925. Washington, DC: Government Printing Office, **1997**
9. Boivin, D., Gendron, S., Beaulieu, E., Gingras, D., Beliveau, R. The antiangiogenic agent Neovastat (AE-941) induces endothelial cell apoptosis. *Mol Cancer Ther.* **2002**, *1*, 795-802.



10. Beliveau, R., Gingras, D., Kruger, E., Lamy, S., Sirois, P., Simard, B., Sirois, M.G., Tranqui, L., Baffert, F., Beaulieu, E., Dimitriadou, V., Pepin, M.-C., Courjal, F., Ricard, I., Poyet, P., Falardeau, P., Figg, W.D, Dupont, E. The antiangiogenic agent neovastat (AE-941) inhibits vascular endothelial growth factor-mediated biological effects. *Clin Cancer Res.* **2002**, *8*, 1242-1250.
11. Sava, G., Ceschia, V., Pacor, S. Mechanism of the antineoplastic action of lysozyme: evidence for host mediated effects. *Anticancer Res.* **1989**, *9*, 1175-1180.
12. Chen, S.F., Fei, X., Li, S.H. A new simple method for the isolation of microvascular endothelial cells. *Microvasc. Res.* **1995**, *50*, 119-128.
13. DeClerck, Y.A., and Laug, W.E. Inhibition of tumor cell collagenolytic activity bovine endothelial cells. *Cancer Res.* **1986**, *46*, 3580-3586.
14. Schagger, H., and von Jagow, G. Tricine-sodium dodecyl sulfate-polyacrylamide gel electrophoresis for the separation of proteins in the range 1 to 100 KDa. *Anal. Biochem.* **1987**, *166*, 368-379.
15. Langer, R., and Folkman, J. Polymer for sustained release of proteins and other macromolecules. *Nature* **1976**, *263*, 797-800.
16. Taylor, S., and Folkman, J. Protamine is an inhibitor of angiogenesis. *Nature* **1982**, *297*, 307-312.

*SamplesAvailability:* Available from the authors.

© 2004 by MDPI (<http://www.mdpi.org>). Reproduction is permitted for noncommercial purposes.