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Communication

# Two New Constituents from Artemisia capillaris Thunb.

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**Abstract:** Two new compounds, 6'-O-caffeoyl-*p*-hydroxyacetophenone-4-O- $\beta$ -D-glucopyranoside (1) and 6-amino-9-[1-(3,4-dihydroxyphenyl)ethyl]-9*H*-purine (2) were isolated from the aerial parts of *Artemisia capillaris* Thunb. The structures were established on the basis of spectral data.

Keywords: Artemisia capillaris; p-hydroxyacetophenone; adenine

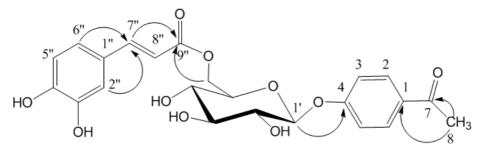
#### Introduction

Infusions of the buds, stems and leaves of *Artemisia capillaris* (Yin Chen Hao, Capillary or Oriental Wormwood) have been used in Chinese Traditional Medicine since antiquity as a cholagogic, antipyretic, anti-inflammatory and diuretic purposes and in the treatment of jaundice [1-6]. Coumarins, flavonol glycosides and a group of unidentified aglycones have been reported so far from the inflorescence of *Artemisia capillaris* [7,8]. Our investigation of the aerial parts of this plant has led to the isolation of two new constituents. This paper deals with the isolation and structural determination of these compounds.

#### **Results and Discussion**

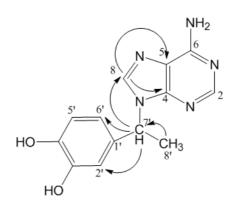
Compound 1, a yellow powder, was formulated as  $C_{23}H_{24}O_{10}$  on the basis of HR-FAB-MS data  $(m/z \ 461.1436, \ [M+H]^+, \ calcd. \ 461.1446)$ . Its <sup>1</sup>H-NMR spectrum indicated one methyl ( $\delta \ 2.38, \ s$ ), a 1,4-disubstituted benzene ring [ $\delta$  7.85 (2H, d, J = 8.7 Hz) and  $\delta$  7.10 (2H, d, J = 8.7 Hz)] and a 1,2,4trisubstituted benzene ring [ $\delta$  6.76 (1H, d, J = 8.1 Hz),  $\delta$  6.96 (1H, d, J = 8.1 Hz) and  $\delta$  7.05 (1H, s)]. In addition, the coupling constants of the proton signals at  $\delta$  7.46 and  $\delta$  6.25 were 15.8 Hz, indicating the presence of two *trans*-olefinic protons. Thus, it was presumed that **1** contained an *O*-caffeoyl group [9,10]. The <sup>1</sup>H-NMR and <sup>13</sup>C-NMR spectra also indicated the presence of a glucopyranosyl unit [ $\delta_{\rm H}$ 5.07 (anomeric proton),  $\delta_{\rm C}$  99.6, 76.4, 74.0, 73.2, 70.1, 63.3]. The coupling constant of the anomeric proton signal (J = 9.0 Hz) indicated the  $\beta$ -configuration of the glucose. Long-range correlations were observed in the heteronuclear multiple-bond connectivity (HMBC) spectrum as follows: (a)  $\delta_H$  7.05 and  $\delta_C$  olefinic carbon 145.8,  $\delta_H$  6.96 and  $\delta_C$  145.8,  $\delta_H$  7.46 and ester carbonyl carbon  $\delta_C$  166.4 which further evidenced the presence of O-caffeoyl group; (b) methyl proton at  $\delta_H$  2.30 and  $\delta_C$  196.4,  $\delta_H$  2.30 and 130.9 indicating that the methyl and 1,4-disubstituted benzene ring was connected by a carbonyl carbon [4]; (c) anomeric proton of glucose  $\delta_H$  5.07 and  $\delta_C$  160.9, H-6' of glucose and ester carbonyl carbon  $\delta_{\rm C}$  166.4 showed that the O-caffeoyl group and 1,4-disubstituted benzene ring must be linked at the position C-6' and C-1' of the glucose, respectively. Acid hydrolysis of 1 furnished the sugar component, which was confirmed as glucose by TLC comparison with an authentic sample. Thus, compound 1 was established to be 6'-O-caffeoyl-p-hydroxyacetophenone-4-O- $\beta$ -D-glucopyranoside (Figure 1).





Compound **2** was obtained as an amorphous powder. The molecular formula was established to be  $C_{13}H_{13}N_5O_2$  based on HR-FAB-MS data (*m/z* 272.1136, [M+H]<sup>+</sup>, calcd for  $C_{13}H_{14}N_5O_2$ : 272.1128). The signals in the <sup>1</sup>H-NMR spectrum of compound **2** at  $\delta$  8.27 (s), 8.11 (s) and 7.19 (2H, s, NH<sub>2</sub>) along with those in the <sup>13</sup>C-NMR spectrum at  $\delta$  118.9, 139.2, 149.2, 152.4 and 156.0 indicated the presence of an adenine ring. The <sup>1</sup>H-NMR spectrum also showed a 1,2,4-trisubstituted benzene ring [ $\delta$  6.67, 1H, d, *J* = 8.4 Hz),  $\delta$  6.64 (1H, d, *J* = 8.4 Hz) and  $\delta$  6.69 (1H, s)] and a methyl which was connected to a methine [ $\delta$  1.85, d, *J* = 7.2 Hz),  $\delta$  5.64 (q, *J* = 7.2 Hz). In its HMBC spectrum long-range correlations were observed between  $\delta_H$  5.64 and  $\delta_C$  139.2 and  $\delta_H$  5.64 and  $\delta_C$  114.0, indicating that both the adenine ring and benzene ring were connected to the methine. In summary, based on HSQC and HMBC spectral data, compound **2** was determined to be 6-amino-9-[1-(3,4-dihydroxyphenyl)ethyl]-9*H*-purine (Figure 2).

#### Figure 2. Important HMBC correlations of 2.



#### **Experimental**

#### General

The NMR data were obtained on a Bruker ARX-300 spectrometer (300 MHz for <sup>1</sup>H and 75 MHz for <sup>13</sup>C) in DMSO-*d6* with TMS as internal standard. The HR-FAB-MS data were obtained on the Micross Mass Autospec-UltimaE TOF mass spectrophotometer. Chromatography was performed on silica gel (200-300 mesh), Sephadex LH-20 and with a Shimadzu LC-8A HPLC instrument equipped with a reversed-phase column.

#### Plant material

The material investigated were aerial parts of *A. capillaris* purchased from the Cooperative of Traditional Chinese Medicine of Shenyang, P.R. China. A voucher specimen was identified by Prof. Qi-Shi Sun and deposited at the School of Traditional Chinese Medicine of Shenyang Pharmaceutical University, P.R.China.

#### Extraction, isolation and product characterization

The dry aerial parts (10 kg) of *A. capillaries* were extracted with boiling water three times, and then precipitated with 75% aqueous alcohol. After evaporation of the solvents under reduced pressure, the residue (1.1 kg) was suspended in H<sub>2</sub>O and extracted sequentially with petroleum ether, ethyl acetate and *n*-butanol. The ethyl acetate extract (53 g) was subjected to silica gel CC with elution by CHCl<sub>3</sub>-CH<sub>3</sub>OH in increasing polarity to obtain eight fractions (A to G). Fraction F was then purified by Sephadex LH-20 column chromatography eluted with CH<sub>3</sub>OH, and further separated by preparative RP-HPLC eluted with 35% aqueous CH<sub>3</sub>OH to give compounds **1** (20 mg) and **2** (8 mg). <sup>1</sup>H- and <sup>13</sup>C-NMR data are shown in Table 1.

Position	1		2	
	Н	С	Н	С
1		130.9		
2	7.84 (d, <i>J</i> =8.7 Hz)	130.2	8.11 (s)	152.4
3	7.10 (d, <i>J</i> =8.7 Hz)	116.0		
4		160.9		149.2
5	7.10 (d, <i>J</i> =8.7 Hz)	116.0		118.9
6	7.84 (d, <i>J</i> =8.7Hz)	130.2		156.0
7		196.4		
8	2.38 (s)	26.3	8.27 (s)	139.2
1'	5.07 (d, <i>J</i> =9.0 Hz)	99.6		132.5
2'	3.30 (m)	73.2	6.69 (s)	114.0
3'	3.32 (m)	76.4		145.2
4'	3.20 (m)	70.1		149.2
5'	3.70 (m)	74.0	6.67 (d, <i>J</i> =8.4 Hz)	115.3
6′	4.41(d, <i>J</i> =11 Hz),	63.3	6.64 (d, <i>J</i> =8.4 Hz)	117.0
	4.17 (dd, <i>J</i> =11, 7.3 Hz)			
7'			5.64 (q, <i>J</i> =7.2 Hz)	52.0
8′			1.85 (d, <i>J</i> =7.2 Hz)	20.6
1″		125.0		
2″	7.05 (s)	114.1		
3″		145.2		
4″		148.8		
5″	6.76 (d, <i>J</i> =8.1 Hz)	115.2		
6″	6.96 (d, <i>J</i> =8.1 Hz)	121.0		
7″	7.46 (d, <i>J</i> =15.8 Hz)	145.8		
8″	6.25 (d, <i>J</i> =15.8 Hz)	113.0		
9″		166.4		

Table 1. NMR data of compounds 1, 2 (ppm from TMS, in DMSO-*d*<sub>6</sub>).

#### Acid Hydrolysis of 1 [12]

Compound **1** (8 mg) was refluxed with 10% HCl in 75% EtOH (3 mL) for 6 h. After cooling, the reaction mixture was extracted with EtOAc (3 ml). The water layers were concentrated and compared with *D*-glucose by TLC analysis [system 1: silica-gel, *n*-BuOH-C<sub>5</sub>H<sub>5</sub>N-H<sub>2</sub>O (8.0:4.0:3.0), *Rf*: 0.24; system 2: silica-gel, EtOAc-MeOH-H<sub>2</sub>O-AcOH (6.5:2.0:1.5:1.5), *Rf*: 0.42]. Those confirmed that the sugar moiety obtained from aqueous acid hydrolysis of compound **1** was *D*-glucose.

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### References

- 1. Tang, W.; Eisenbrand, G. Chinese Drugs of Plant Origin, Chemistry, Phamacology and use in traditional and modern medicine. Springer Verlag: New York, **1992**; p. 179.
- 2. Han, K.-H.; Jeon, Y.-J.; Athukorala, Y.; Choi, K.-D.; Kim, C.-J.; Cho, J.-K.; Sekikawa, M.; Lee, C.-H. A water extract of Artemisia capillaris prevents 2,2'-azobis(2-amidinopropane) dihydrochloride-induced liver damage in rats. *J. Med. Food* **2006**, *9*, 342-347.
- 3. Han, J.; Zhao, Y.-L.; Shan, Li-M.; Huang, F.-J.; Xiao, X.-H. An experiment on standardized cell culture assay in assessing the activities of Composite Artemisia Capillaris Tablets against hepatitis B virus replication in vitro. *Chin. J. Integr. Med.* **2005**, *11*, 54-56.
- 4. Jang, S.; Kim, Y.-J.; Lee, W.-Y.; Kwak, K. C.; Baek, S. H.; Kwak, G. B.; Yun, Y.-G.; Chai, K.-Y. Scoparone from Artemisia capillaris inhibits the release of inflammatory mediators in RAW 264.7 cells upon stimulation cells by interferon-gamma Plus LPS. *Arch. Pharm. Res.* **2005**, *28*, 203-208.
- 5. Hong, S. H.; Seo, S. H.; Lee, J. H.; Choi, B. T. The aqueous extract from Artemisia capillaris Thunb. inhibits lipopolysaccharide-induced inflammatory response through preventing NFkappaB activation in human hepatoma cell line and rat liver. *Int. J. Mol. Med.* **2004**, *13*, 717-720.
- 6. Hu, Y. Q.; Tan, R. X.; Chu, M. Y.; Zhou, J. Apoptosis in human hepatoma cell line SMMC-7721 induced by water-soluble macromolecular components of Artemisia capillaris Thunberg. *Jap. J. Cancer Res.: Gann* **2000**, *91*, 113-117.
- 7. Yamahara, J.; Kobayashi, G.; Matsuda, H.; Katayama, T.; Fujimura, H. The effect of scoparone, a coumarin derivative isolated from the Chinese crude drug *Artemisiae capillaris flos*, on the heart. *Chem. Pharm. Bull.* **1989**, *37*, 1279-1299.
- 8. Fakeya, K.; Yoshitomo, N.; Haruji, O. Studies on 'Inchinko' II. Studies on the compounds related to capillarisin and flavonoids. *Yakugaku Zasshi* **1976**, *96*, 855-862.
- Yang, Z. G.; Li, H. R.; Wang, L. Y.; Li, Y. H.; Lu, S. G.; Wen, X. F.; Wang, J.; Akihiro, D.; Susumu, K. Triterpenoids from *Hippophae rhamnoides* L. and Their Nitric OxideProduction-Inhibitory and DPPH Radical-Scavenging Activities. *Chem. Pharm. Bull.* 2007, 55, 15-18.
- Logendra, S.; Ribnicky, D. M.; Yang, H.; Poulev, A.; Ma, J.; Kennelly, E. J.; Raskin, I. Bioassayguided isolation of aldose reductase inhibitors from Artemisia dracunculus. *Phytochemistry* 2006, 67, 1539-1546.
- 11. Shao, Y.; Li, Y. L; Zhou, B. N. Structural elucidation and synthesis of asterbatanoside A from *Aster batanfensis. Chin. Chem. Lett.* **1994**, *5*, 675-678.
- 12. Sun, J.-M.; Yang, J.-S.; Zhang, H. Two New Flavanone Glycosides of *Jasminum lanceolarium* and Their Anti-oxidant Activities. *Chem. Pharm. Bull.* **2007**, *55*, 474-476.

Sample Availability: Available from the authors.

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